

Determining the distribution of platypuses & shortfinned eels in the Moorabool River

Prepared for: Jackson Cass **Prepared by:** Josh Griffiths, Sarah Licul and Dr Andrew Weeks

Moorabool Catchment Landcare Group PO Box 239 Ballan, VIC 3342 Australia EnviroDNA 293 Royal Parade Parkville, VIC 3052 Australia

Disclaimer

The professional analysis and advice in this report has been prepared for the exclusive use of the party or parties to whom it is addressed (the addressee) and for the purposes specified in it. This report is supplied in good faith and reflects the knowledge, expertise and experience of the consultants involved. The report must not be published, quoted or disseminated to any other party without prior written consent from EnviroDNA pty ltd.

EnviroDNA pty ltd accepts no responsibility whatsoever for any loss occasioned by any person acting or refraining from action as a result of reliance on the report. In conducting the analysis in this report EnviroDNA pty ltd has endeavoured to use what it considers is the best information available at the date of publication including information supplied by the addressee. Unless stated otherwise EnviroDNA pty ltd does not warrant the accuracy of any forecast or prediction in this report.



Project team

Title	Name
Project Manager	Josh Griffiths
Project Consultant	Sarah Licul
Project Supervisor	Dr. Andrew Weeks

Version control

Date	Version	Description	Author	Reviewed By
27/06/2021	1.0	Final Report_Draft	JG	JC
30/06/21	2.0	Final Report	JG	

Abbreviations

Abbreviations	Description
eDNA	environmental DNA
MCLG	Moorabool Catchment Landcare Group
CCMA	Corangamite Catchment Management Authority
ISC	Index of Stream Condition 2010
qPCR	quantitative Polymerase Chain Reaction





Contents

Background	. 4
Methods	. 5
Investigating platypus distribution Assessing condition of waterways	. 5 . 7
Findings	. 8
Current distribution of platypuses Current distribution of shortfin eels	. 8 . 8
Condition of waterways	. 8
Discussion and recommendations	11
Acknowledgments	13
References	14
Appendix 1. Site locations, eDNA results, river health assessments and ISC scores	18
Appendix 2. Site photos representing different river health conditions	19



Background

The platypus (*Ornithorhynchus anatinus*) is a semi-aquatic mammal that inhabits a variety of freshwater habitats throughout eastern Australia (Grant 1992). The species was recently listed as "Near Threatened" by the IUCN (Woinarski *et al.* 2014; Woinarski and Burbidge 2016) and "Vulnerable" in Victoria in recognition of mounting evidence of population declines and localised extinctions throughout its range (Bino *et al.* 2020). Difficulties in assessing platypus populations and lack of historical data have hampered studies to quantify the impacts of various threats. However, the platypus populations are vulnerable to a number of potential threats including drought, altered flow regimes, changes to surrounding catchment area due to agriculture or urbanisation, removal of riparian vegetation, habitat fragmentation, poor water quality, and predation from invasive predators (Grant and Temple-Smith 1998, 2003).

The Moorabool Catchment has been significantly modified due to widespread land clearing for agriculture, presence of invasive species (e.g. willows), poor water quality, and reduced river flows due to water extraction and diversion as well as climate change. The Moorabool River is estimated to receive only 10% of its natural flow (ref) and was rated as Moderate to Very Poor by the last Index of Stream Condition assessment in 2010 (Department of Environment and Primary Industries 2010). All of these factors can be expected to have impacts on platypus populations (and other aquatic dependent species), however little contemporary information exists on the current distribution or abundance of platypuses in the area. Platypus populations in other similar areas in Victoria (i.e. agricultural areas of the Wimmera, Corangamite and Campaspe Catchments) are considered under serious stress although quantifying declines is limited by lack of rigorous historical data (Serena *et al.* 2002; Griffiths *et al.* 2018; Griffiths and Weeks 2018; Griffiths *et al.* 2019; Griffiths and Licul 2020)

This project aimed to address the lack of data by using environmental DNA to investigate the current distribution of platypuses throughout the Moorabool Catchment while engaging the local community in conservation issues. Available data is limited to anecdotal reports of platypus sightings and no systematic surveys have previously been undertaken. Environmental DNA (eDNA) is a non-invasive sampling technique that detects genetic material from a target species secreted into its surrounding environment (water). Quantitative comparisons with traditional sampling methods already indicate that eDNA methods are superior in terms of sensitivity and cost efficiency, particularly for scarce, elusive or cryptic species (Biggs *et al.* 2015; Smart *et al.* 2015), including platypuses (Lugg *et al.* 2018; Weeks *et al.* 2015), enabling effective detection at low densities. As part of the project, we also used the eDNA samples to investigate the distribution of short-finned eels (*Anguilla australis*). The project aimed to identify declines where possible, provide a comprehensive baseline for future monitoring and direct management actions to improve the long term viability of platypus populations.



Methods

Investigating platypus distribution.

Eighteen survey sites were selected in collaboration with Moorabool Catchment Landcare Group (MCLG) to include a variety of waterways and habitat and provide good spatial coverage throughout the region (Figure 1, Appendix 1). Sampling sites were selected with consideration of known previous distribution of platypuses, recent sightings, and accessibility of sites along the waterways. Historical data on platypus distribution was collated from online databases (www.ala.org.au, www.vba.vic.gov.au, www.platypusSPOT.org).

The current occurrence of platypuses and short-fin eel at each site was determined using environmental DNA techniques. Water sampling was undertaken during May 2021 to target the juvenile dispersal period for platypuses in Victoria (Grant 2007) and ensure adequate surface water availability. Water samples were collected by volunteers from the MCLG and local residents following detailed instructions and demonstration of correct sampling techniques by EnviroDNA. At each site, water samples were collected in duplicate by passing up to 400 ml water (average 200 ml) through a 0.22 μ m filter (Sterivex). Filtration was undertaken on site to reduce DNA degradation during transport of whole water samples (Yamanaka et al. 2016). Clean sampling protocols were employed to minimise contamination including new sampling equipment at each site, not entering water, and taking care not to transfer soil, water or vegetation between sites. Filters were stored on ice for a maximum of 48 hrs before being transported to the laboratory for processing.

DNA was extracted from the filters using a commercially available DNA extraction kit (Qiagen DNeasy Blood and Tissue Kit). Real-time quantitative Polymerase Chain Reaction (qPCR) assays were used to amplify the target DNA, using species-specific markers targeting a small region of the mitochondrial DNA, previously developed and assessed for specificity and sensitivity by EnviroDNA (e.g. Lugg *et al.* 2018; Weeks *et al.* 2015). Assays were performed in triplicate on each sample. Negative controls were included for the DNA extraction and qPCR steps. At least two positive PCR's (out of six or nine assays undertaken for the site) were required to classify the site as positive for the presence of platypus.





Figure 1. Location of sampling sites for eDNA analysis.

Page 6 Project number 1904CR1 EnviroDNA Pty Ltd +61 3 9028 8753 envirodna.com



Assessing condition of waterways.

River health was assessed at each site by volunteers based on an established methodology previously used by Victorian CMA's and WaterWatch that incorporates in-stream and riparian variables (River Detectives www.riverdetectives.net.au). An additional platypus-specific variable was added to rate availability of burrowing habitat. The river health assessment was explained and demonstrated to the volunteer group during the training session and habitat variables related to platypus requirements to provide context. The six habitat variables are evaluated and assigned to five categories from Very Poor to Excellent. These are then tallied to provide an overall river health rating for the site.

A measure of river health at the reach level was also derived from 2010 Index of Stream Condition (ISC) scores (Department of Environment and Primary Industries 2010) to support volunteer assessments for the waterway reaches where ISC assessments had taken place. ISC provides an overall measure of river health (very poor, poor, moderate, good, excellent) based on five key metrics: hydrology, streamside zone, physical form, water quality, and aquatic life.





Findings

Current distribution of platypuses.

Six of the 18 sites sampled returned positive results for platypus DNA (33%; Figure 2, Appendix 1). Trace amounts of DNA was detected at four other sites but was not above the defined threshold level (at least 2 positive PCR's) to be considered positive (indicated as equivocal in Appendix 1). While equivocal results may indicate the species presence at very low abundance, it can also arise from field contamination during sampling or dispersal of DNA from further upstream. Repeat sampling is recommended to confirm presence or absence at these sites. Detections of platypus DNA was clustered in sites around and downstream of the junction of the east and west branches of the Moorabool River although their distribution may also extend up the east branch in low abundance (Figure 2).

Current distribution of shortfin eels.

Four of the 18 sites sampled returned positive results for shortfin eel DNA (22%, Figure 3, Appendix 1). Trace amounts of DNA was detected at four other sites but was not above the defined threshold level (at least 2 positive PCR's) to be considered positive (indicated as equivocal in Appendix 1). Positive detections of shortfin eel DNA were recorded in the upper reaches of both east and west branches of the Moorabool River with equivocal results indicating their distribution may extend further downstream at low abundance (Figure 3).

Condition of waterways

River health assessments were completed at 15 sites. The sites were assessed as Degraded (1), Poor (2), Fair (3), Good (7), or Excellent (2) (Appendix 2). Fifteen sampling sites were located in reaches that also had corresponding ISC 2010 scores that were rated as poor (3) or fair (12)(Appendix 1). Detections of platypus eDNA tended to occur at sites with better health ratings by both volunteers or ISC although sample sizes were too small for analysis.





Figure 2. Results from the eDNA sampling indicating positive detection (green), equivocal (pale green) or non-detection (grey) of platypus DNA and historical platypus records from online databases (red <10yrs, yellow >10yrs).

Page 9 Project number 1904CR1 EnviroDNA Pty Ltd +61 3 9028 8753 envirodna.com





Figure 3. Results from the eDNA sampling indicating positive detection (red), equivocal (pink) or non-detection (grey) of shortfin eel DNA.

Page 10 Project number 1904CR1 EnviroDNA Pty Ltd +61 3 9028 8753 envirodna.com



Discussion and recommendations

There is very limited recent or historical data on platypuses in the Moorabool Catchment with just 16 records scattered throughout the study area between 1994 to 2019. Records from online wildlife databases (Atlas of Living Australia, Victorian Biodiversity Atlas, platypusSPOT) mostly comprise anecdotal sightings from residents or visitors and typically are concentrated near population centres or tourist locations. Despite these limitations, the data indicate platypuses were likely to have been widespread throughout the Moorabool River and its perennial tributaries as well as the upper Werribee River as far as reliable water is available. Results from the current study broadly correspond with these previous records apart from the upper Werribee River where no platypus eDNA was detected from two sampling sites. However, positive eDNA results have been previously recorded at Ballan (2019, EnviroDNA unpublished data) as well as recent sightings and low captures from live-trapping surveys (Griffiths *et al.* 2012; Griffiths, Kelly, van Rooyen, *et al.* 2014) all suggesting platypuses occur in low abundance in the upper Werribee River.

In the absence of any previous systematic data, it is impossible to infer a population trajectory. The current results indicate platypuses are widely distributed throughout the Moorabool River including the east and west branches although there was no evidence of platypuses upstream of Bostock Reservoir. Poor habitat quality in Paddock Creek and the barrier posed by the reservoir may prevent platypus dispersal into this area. Positive eDNA results have also been recorded downstream of the current study area (between Meredith and Bannockburn, EnviroDNA unpublished data) indicating platypuses likely occupy the lower Moorabool River downstream to the junction with the Barwon River near Geelong.

Although widely distributed, the low site occupancy (33% although it could be as high as 56% if equivocal results are true positives) indicating platypuses are likely to be in relatively low abundance throughout the area. Similar results have been recorded in the adjacent upper Barwon region (23-31%; Griffiths *et al.* 2019). This likely reflects poor habitat quality with ISC ratings for the river reaches considered poor to fair. River health assessments at the sampling sites were generally higher than the ISC ratings (Appendix 2). Differences in timing, scale, and metrics between the two methods likely account for some of the discrepancies. The last ISC assessment was undertaken in 2010 at the end of an extensive period of drought and conditions may have changed since then and the current study in 2021. The ISC assessed condition at the reach scale which is usually several kilometres (or tens of kilometres) long while volunteers in this study assessed the visible area at each site (approximately 50 m). It is possible for small patches of relatively good habitat to exist along poor reaches (or vice versa). The ISC also incorporates several metrics such as hydrology and aquatic life (invertebrates) that are unable to be easily assessed by untrained volunteers in the field.

Results from the current study as well as previous eDNA testing indicate the middle reaches of the Moorabool River is the stronghold for platypuses in the region. Not surprisingly, this also corresponds with somewhat better habitat quality as rated by both ISC and our volunteers. Habitat variables known to be important for platypuses include large riparian trees, overhanging vegetation, pools 1-3 m deep, near vertical, undercut and stabilised banks at least 0.5 m above the water, large woody debris, and coarse benthic substrates (Bethge *et al.* 2003; Ellem *et al.* 1998; Grant 2004; M. Serena *et al.* 1998; Serena *et al.*

EnviroDNA Pty Ltd +61 3 9028 8753 envirodna.com



2001; Worley and Serena 2000; M Serena *et al.* 1998). Critically, platypuses require aquatic habitats that support adequate and reliable resources of macroinvertebrate prey and many of the habitat variables above provide shelter and food resources for aquatic macroinvertebrates. Waterways with reduced baseflows, and/or high flow variability, or poor water quality such as high turbidity (i.e. sedimentation arising from degraded riparian zones and subsequent erosion) or low dissolved oxygen can result in depauperate macroinvertebrate assemblages (Chessman 2009; Marchant and Grant 2015; Boulton and Brock 1999; Walsh and Webb 2013).

Like platypuses, shortfin eels also have fairly generic habitat requirements, although eels tend to prefer slower flowing or still waters. Critically for eels as a diadromous species, they require hydrologic connections to the marine environment for spawning migrations. The hydrologic score of the ISC in the lower Moorabool River was very poor. Poor flow regimes, including cease to flow events could limit dispersal of eels to the ocean as well as limit habitat suitability for platypuses and connection with the lower Barwon population. While flows may have improved in the last 10 years, ensuring adequate baseflows throughout the Moorabool system is critical to maintaining river health and habitat quality for a variety of aquatic species.

It is important to note that results from the current study were obtained from a single sampling event and represent a snapshot of platypus and shortfin eel distribution at the time of sampling only. Previous studies have demonstrated that eDNA has high sensitivity to detect platypuses, even at low densities (Lugg et al. 2018; Weeks et al. 2015), but negative site results may still arise in waterways where platypuses are known to occur if no platypuses have been active near the sampling site. In freshwater systems, eDNA generally degrades or disperses relatively quickly (i.e. within days) (Thomsen et al. 2012; Pilliod et al. 2014). In addition, platypuses are highly mobile with typical home ranges of several kilometers (Gardner and Serena 1995; Serena and Williams 2012; Grant et al. 1992; M. Serena et al. 1998). Species behaviour, movements, and habitat use can also change in response to seasons and environmental conditions (Gust and Handasyde 1995; Griffiths and Weeks 2015; Griffiths, Kelly and Weeks 2014). Therefore, some temporal variation in localised occurrence of platypuses and eels is expected although broad distribution should remain similar over short time periods. Therefore, these results provide a good indication of the target species' distribution throughout the area. Importantly, there is now baseline data against which to assess future changes in platypus or eel populations in response to natural disturbances or management actions.



Acknowledgments

EnviroDNA would like to acknowledge MCLG, CCMA, and Landcare Australia for supporting and funding this project. Special thanks to Jennifer Johnson and Jackson Cass of MCLG for coordinating and organising such a successful citizen science day and all the MCLG members and local community for attending and collecting samples.



References

Bethge P., Munks S., Otley H. & Nicol S. (2003) Diving behaviour, dive cycles and aerobic dive limit in the platypus *Ornithorhynchus anatinus*. *Comp. Biochem. Physiol. - A Mol. Integr. Physiol.* **136**, 799–809.

Biggs J., Ewald N., Valentini A. *et al.* (2015) Using eDNA to develop a national citizen science-based monitoring programme for the great crested newt (*Triturus cristatus*). *Biol. Conserv.* doi: 10.1016/j.biocon.2014.11.029. [online].

Bino G., Kingsford R. T. & Wintle B. A. (2020) A stitch in time – Synergistic impacts to platypus metapopulation extinction risk. *Biol. Conserv.* **242**, 108399. [online].

Boulton A. J. & Brock M. A. (1999) *Australian Freshwater Ecology: processes and management.* Gleneagles Publishing, Adelaide.

Chessman B. C. (2009) Climatic changes and 13-year trends in stream macroinvertebrate assemblages in New South Wales, Australia. *Glob. Chang. Biol.* **15**, 2791–2802.

Department of Environment and Primary Industries (2010) *Index of Stream Condition. The third benchmark of Victorian river condition.* Melbourne.

Ellem B. A., Bryant A. & O'Connor A. (1998) Statistical modelling of platypus, *Ornithorhynchus anatinus*, habitat preferences using generalized linear models. *Aust. Mammal.*

Frankham R., Ballou J. D. & Briscoe D. A. (2010) *Introduction to conservation genetics*. 2nd edn. Cambridge University Press, Cambridge, UK.

Gardner J. L. & Serena M. (1995) Spatial organization and movement patterns of adult male platypus, *Ornithorhynchus anatinus* (monotremata: Ornithorhynchidae). *Aust. J. Zool.* **43**, 91–103.

Grant T. (2004) Depth and substrate selection by platypuses, *Ornithorhynchus anatinus*, in the lower Hastings River, New South Wales. *Proc. Linn. Soc. New South Wales*.

Grant T. R. (1992) Historical and current distribution of the platypus, *Ornithorhynchus anatinus*, in Australia. In: *Platypus and Echidnas* (ed M. L. Augee) pp. 232–254 Royal Zoological Society of New South Wales.

Grant T. R. (2007) Platypus. 4th ed. CSIRO publishing: Australian natural history series.

Grant T. R., Grigg G. C., Beard L. A. & Augee M. L. (1992) Movements and burrow use by platypuses, *Ornithorhynchus anatinus*, in the Thredbo River, New South Wales. In: *Platypus and Echidnas* (ed M. L. Augee) pp. 263–267 Royal Zoological Society of New South Wales. [online].





Grant T. R. & Temple-Smith P. D. (1998) Field biology of the platypus (*Ornithorhynchus anatinus*): historical and current perspectives. *Philos. Trans. R. Soc. London Ser. B-Biological Sci.* **353**, 1081–1091.

Grant T. R. & Temple-Smith P. D. (2003) Conservation of the platypus, *Ornithorhynchus anatinus*: Threats and challenges. *Aquat. Ecosyst. Heal. Manag.* **6**, 5–18.

Griffiths J., Kelly T., van Rooyen A. & Weeks A. (2014) *Distribution and relative abundance of platypuses in the greater Melbourne area: survey results 2013/14. (Report to Melbourne Water).* cesar, Parkville.

Griffiths J., Kelly T. & Weeks A. (2012) *Distribution and relative abundance of platypuses in the greater Melbourne area: survey results 2011/12. (Report to Melbourne Water).* cesar, Parkville.

Griffiths J., Kelly T. & Weeks A. (2014) *Impacts of high flows on platypus movements and habitat use in an urban stream. (Report to Melbourne Water).* **cesar**, Parkville, VIC.

Griffiths J. & Licul S. (2020) Mapping the distribution of platypuses and blackfish in the upper Campaspe region using citizen scientists and environmental DNA. Prepared for: Upper Campaspe Landcare Network. EnviroDNA, Parkville, VIC.

Griffiths J., Song S. & Weeks A. (2019) *Determining the distribution of platypuses in the upper Barwon region using environmental DNA. (Prepared for Upper Barwon Landcare Network)*. EnviroDNA, Parkville, VIC.

Griffiths J., Song S. & Weeks A. R. (2018) *Platypus distribution , relative abundance and genetic health in the MacKenzie River 2018 (Report to Wimmera CMA).* cesar, Parkville, VIC.

Griffiths J. & Weeks A. (2015) Impact of environmental flows on platypuses in a regulated river. Report to Melbourne Water. Parkville, VIC.

Griffiths J. & Weeks A. (2018) Investigating the current distribution of platypuses, rakali, and blackfish in the upper Wimmera region (Report to Project Platypus). EnviroDNA, Parkville, VIC.

Gust N. & Handasyde K. (1995) Seasonal variation in the ranging behaviour of the platypus (*Ornithorhynchus anatinus*) on the Goulburn River, Victoria. *Aust. J. Zool.* **43** , 193–208.

Lugg W. H., Griffiths J., van Rooyen A. R., Weeks A. R. & Tingley R. (2018) Optimal survey designs for environmental DNA sampling. *Methods Ecol. Evol.* **9**, 1049–1059.

Marchant R. & Grant T. R. (2015) The productivity of the macroinvertebrate prey of the platypus in the upper Shoalhaven River, New South Wales. *Mar. Freshw. Res.* **66**, 1128–1137.

Pilliod D. S., Goldberg C. S., Arkle R. S. & Waits L. P. (2014) Factors influencing detection of eDNA from a stream-dwelling amphibian. *Mol. Ecol. Resour.*



Serena M, Thomas J. L. & Williams G. A. (1998) Status and habitat relationships of platypus in the Dandenong Creek Catchment: II. Results of surveys and radio-tracking studies, September 1997 - March 1998. (Report to Melbourne Water). Australian Platypus Conservancy, Whittlesea.

Serena M., Thomas J. L., Williams G. A. & Officer R. C. E. (1998) Use of stream and river habitats by the platypus, *Ornithorhynchus anatinus*, in an urban fringe environment. *Aust. J. Zool.* **46**, 267–282.

Serena M. & Williams G. A. (2012) Movements and cumulative range size of the platypus (Ornithorhynchus anatinus) inferred from mark-recapture studies. *Aust. J. Zool.* **60**, 352–359.

Serena M., Williams G. A. & Johnston L. D. (2002) *Status of platypus in the Curdies River catchment: live-trapping and local sightings, summer 2002. (Report to Corangamite Catchment Management Authority).* Australian Platypus Conservancy, Whittlesea.

Serena M., Worley M., Swinnerton M. & Williams G. A. (2001) Effect of food availability and habitat on the distribution of platypus (*Ornithorhynchus anatinus*) foraging activity. *Aust. J. Zool.* **49**, 263–277.

Smart A. S., Tingley R., Weeks A. R., Van Rooyen A. R. & McCarthy M. A. (2015) Environmental DNA sampling is more sensitive than a traditional survey technique for detecting an aquatic invader. *Ecol. Appl.* **25**, 1944–1952.

Soule M. E. (1980) Thresholds for survival: maintaining fitnessand evolutionary potential. In: *Conservation Biology: An Evolutionary-Ecological Perspective* (eds M. E. Soule & B. A. Wilcox) pp. 151–169 Sinauer, Sunderland, MA.

Thomsen P. F., Kielgast J., Iversen L. L. *et al.* (2012) Monitoring endangered freshwater biodiversity using environmental DNA. *Mol. Ecol.* **21**, 2565–2573.

Walsh C. J. & Webb J. A. (2013) *The influence of land use on stream macroinvertebrate assemblage condition in the Melbourne Water management area.* Report prepared for Melbourne Water, Melbourne.

Weeks A., van Rooyen A., Griffiths J. & Tingley R. (2015) *Determining the effectiveness of eDNA for monitoring platypuses (Report to Melbourne Water)*. **cesar**, Parkville.

Woinarski J. & Burbidge A. A. (2016) *Ornithorhynchus anatinus*. The IUCN Red List of Threatened Species.

Woinarski J. C. Z., Burbidge A. A. & Harrison P. L. (2014) *The action plan for Australian mammals 2012.* CSIRO Publishing, Collingwood.

Worley M. & Serena M. (2000) *Ecology and conservation of platypuses in the Wimmera River catchment. IV. Results of habitat studies (Report to Rio Tinto Project Platypus).* Australian Platypus Conservancy, Whittlesea. Yamanaka H., Motozawa H., Tsuji S., Miyazawa R. C., Takahara T. & Minamoto T. (2016) On-site filtration of water samples for environmental DNA analysis to avoid DNA degradation during transportation. *Ecol. Res.* **31**, 963–967.



				Platypus		Shortfin eel			
Sito	Watorway	Asterway Latitude Longitude +ve Site		Site	+ve Site		Site condition	ISC reach	
Sile	Waterway	Latitude	Longitude	assays	result	assays	result	rating	score
1	Werribee River	-37.551041	144.198281	0	Negative	0	Negative	Fair	Poor
2	Werribee River	-37.597396	144.230777	0	Negative	0	Negative	Fair	Poor
з	Moorabool River East	-37 620852	144 162233	1	Equivocal	4	Positivo	Fair	
5	Branch	-57.029032	144.102233	1	Lquivocai		i ositive		Fair
Δ	Moorabool River East 2	Positivo							
т	Branch	-37.65309	144.1765	1	Lquivocai		1 OSITIVE		Fair
5	Moorabool River East	-37 742801	144 132492	1	Equivocal	1	Equivocal	Excellent	
Ŭ	Branch	07.142001	144.102432		Equivoodi		Equivoodi		Poor
6	Moorabool River West			0	Negative	2	Positive		
0	Branch	-37.60757	144.19278	Ŭ	lioguaro				Fair
7	Moorabool River West	-37 711349	144 09568	6	Positive	4	Positive	Good	
	Branch			Ŭ					Fair
8	Moorabool River	-37.769292	144.108861	4	Positive	0	Negative	Good	Fair
9	Moorabool River	-37.776527	144.111268	4	Positive	0	Negative	Good	Fair
10	Moorabool River	-37.78871	144.108906	4	Positive	1	Equivocal	Good	Fair
11	Moorabool River	-37.820109	144.106021	3	Positive	0	Negative	Good	Fair
12	Moorabool River	-37.831556	144.118596	3	Positive	1	Equivocal		Fair
13	Moorabool River	-37.832895	144.128392	1	Equivocal	1	Equivocal	Good	Fair
14	Moorabool River	-37.872274	144.130089	0	Negative	0	Negative	Excellent	Fair
15	Paddock Creek	-37.578436	144.104433	0	Negative	0	Negative	Degraded	
16	Paddock Creek	-37.58606	144.12042	0	Negative	0	Negative	Good	
17	Paddock Creek	-37.594752	144.164255	0	Negative	0	Negative	Poor	
18	Bostock Reservoir	-37.600503	144.181392	0	Negative	0	Negative	Poor	Fair

Appendix 1. Sampling site details, eDNA results, river health assessments and ISC scores.

Page 18 Project number 0901CR3



Appendix 2. Site habitat assessments by volunteers.

Sit e	Waterway	Latitude	Longitude	Bank erosion	Bank vegetation	Burrowing habitat	Instream complexity	Channel complexity	Verge vegetation	Site conditio n rating
1	Werribee River	-37.551041	144.19828 1	Fair	Fair	Good	Fair	Excellent	Fair	Fair
2	Werribee River	-37.597396	144.23077 7	Good	Good	Fair	Good	Fair	Poor	Fair
3	Moorabool River East Branch	-37.629852	144.16223 3	Good	Fair	Fair	Poor	Fair	Poor	Fair
4	Moorabool River East Branch	-37.65309	144.1765							
5	Moorabool River East Branch	-37.742801	144.13249 2	Good	Good	Good	Excellent	Excellent	Good	Excellent
6	Moorabool River West Branch	-37.60757	144.19278							
7	Moorabool River West Branch	-37.711349	144.09568	Good	Fair	Fair	Excellent	Excellent	Excellent	Good
8	Moorabool River	-37.769292	144.10886 1	Fair	Fair	Fair	Good	Excellent	Poor	Good
9	Moorabool River	-37.776527	144.11126 8	Excellent	Fair	Fair	Excellent	Good	Fair	Good
10	Moorabool River	-37.78871	144.10891	Good	Good	Good	Fair	Good	Fair	Good
11	Moorabool River	-37.820109	144.10602 1	Excellent	Fair	Excellent	Fair	Fair	Good	Good
12	Moorabool River	-37.831556	144.11859 6							
13	Moorabool River	-37.832895	144.12839	Good	Good	Fair	Excellent	Excellent	Fair	Good

Page 19 Project number 0901CR3 EnviroDNA Pty Ltd +61 3 9349 4723 envirodna.com



			2							
14	Moorabool River	-37.872274	144.13008 9	Excellent	Good	Excellent	Excellent	Excellent	Fair	Excellent
15	Paddock Creek	-37.578436	144.10443 3	Degraded	Degraded	Poor	Degraded	Fair	Fair	Degraded
16	Paddock Creek	-37.58606	144.12042	Excellent	Good	Fair	Fair	Fair	Excellent	Good
17	Paddock Creek	-37.594752	144.16425 5	Degraded	Poor	Poor	Degraded	Poor	Poor	Poor
18	Bostock Reservoir	-37.60050	144.18139 2	Fair	Degraded	Poor	Poor	Poor	Fair	Poor

EnviroDNA Pty Ltd +61 3 9349 4723 envirodna.com





